Physico-Chemical and Antioxidant Properties of Skimmed Varenets (Slavic Baked Milk Yogurt) Mixed with Enzyme-Modified Potato Starches

ELENA V. NIKITINA*, TATIANA A. YURTAEVA, MAXIM S. TSYGANOV and GALINA O. EZHKOVA

Department of Meat and Milk Technology, Faculty of Food Technology, Kazan National Research Technological University, Kazan, Russia.

Abstract
The present studies have examined the effect of potato starches treated with amylase B. licheniformis (Bl) (laboratory sample) or Amylosubtilin® (AM) (Berdsk Factory of Biological Preparations (now: Sibbiofarm), Russia) in different concentrations on the quality of Slavic skim milk drink Varenets made from baked milk. The baked milk is milk heated to 98 °C for 3 hours, it has a sweet aroma and taste and creamy hue. The presence of enzyme-modified potato starches (Bl or AM) has been found to promote the activation of lactic acid fermentation, the accumulation of exopolysaccharides, and such stabilized products have a higher viscosity, a lower percentage of syneresis compared to the control sample. The antioxidant capacity of the Varents samples after fermentation was evaluated by two assays: analysis of radical capture activity of 2,2-diphenyl-1-picrylhydrazyl (DPPH •); and the ability to restore Fe + 3 (Iron Reduction Antioxidant Ability Assay, FRAP). All Varentz samples showed different values for DPPH, FRAP assays depending on the starches used. The Introduction of pre-fermented starches into defatted jam promotes formation of enhanced antioxidant properties of milk product. The use of starches modified with enzymes improves sensory characteristics, in particular, as a fat imitator, formsfull taste of the drink.

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Introduction
Varenets is a traditional dairy product of Slavic (Russian) cuisine, in particular the Urals and Siberia. Very similar drinks can be found in other countries. For example, in Ukraine - ryazhenka, in Georgia such a product is called mazoni, in Egypt - leben, and in Bulgaria and among the Turkic peoples - katyk. For its preparation, traditional starters for yogurt with the bacteria Streptococcus thermophilus and Lactobacillus delbrueckii ssp bulgaricus. are used.

CONTACT
Elena V. Nikitina ev-nikitina@inbox.ru Department of Meat and Milk Technology, Faculty of Food Technology, Kazan National Research Technological University, Kazan, Russia.
A feature of Varenets is the use of melted cow's milk for its preparation. Melted milk is milk heated to 98°C for 3 hours, as a result, the fermented milk product obtained from such milk has a creamy tint and a specific pleasant smell of heating. Traditionally, the fat content of Varenets ranges from 2.5 to 4%, such a drink has a thick consistency, a full saturated taste.

Due to changes in the lifestyle and diet of a large part of the population, the problem of the growth of non-communicable diseases, in particular obesity, diabetes, cardiovascular disease and cancer, is acute. In 2014, the results of a systematic analysis of the dynamics of overweight and obesity in 1980-2013 of adults and children were published, which showed that 39% of adults over 18 years of age were overweight, 13% were obese. The prevalence of obesity among children and adolescents is increasing in both developed and developing countries.

This research reported estimates for 188 countries, 21 regions, and development status (developed or developing). One of the main problems associated with excessive consumption, including fats, is the development of obesity and, as a result, an increased risk of type 2 diabetes, coronary heart disease, hypertension, gallbladder pathologies, some types of cancer, dyslipidemia and other insulin resistance.

This situation contributes to the development of low-calorie, low-fat or low-fat food. Such foods should meet the dietary needs of obese people, people at risk of cardiovascular disease, diabetics and people following a diet for weight loss, and encourage them to buy such products. Here are forecasts that the need for low-fat and low-fat food will steadily increase.

In this regard, it is relevant to obtain the Varenets fermented milk product popular in Russia, the fat content of which is reduced to a minimum value of less than 0.5%. However, the rheological and physical characteristics of defatted dairy products are reduced because the reduction in the proportion of fat changes its mechanical and sensory characteristics. Thermostatic fat-free yogurt has a low hardness and a high degree of syneresis or serum separation on the surface.

One approach that can be implemented in the technology of making Varenets is the use of stabilizers and thickeners of a polysaccharide nature. Carboxymethyl cellulose, gelatin, various starches can be used as stabilizer.

Starch is widely used in the production of yoghurts and other dairy products, their presence improves the texture, gives the defatted product a better and more complete flavor of potato starch is also used for this purpose. The purpose of this work was to identify the influence of enzymatically modified potato starches (EMS) on the physicochemical, rheological, sensory and antioxidant properties of fat-free Varenets.

<table>
<thead>
<tr>
<th>Varenets samples with Starches</th>
<th>Amylase activity (U/g starch)</th>
<th>Amylosubtilin (g/100 ml reaction mixture)</th>
<th>Amylase B. licheniformis (ml/100 ml reaction mixture)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (without starch)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Native (potato starch)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>AM-0.05</td>
<td>0.415</td>
<td>0.01</td>
<td>-</td>
</tr>
<tr>
<td>AM-0.1</td>
<td>0.83</td>
<td>0.0201</td>
<td>-</td>
</tr>
<tr>
<td>AM-0.5</td>
<td>4.15</td>
<td>0.1005</td>
<td>-</td>
</tr>
<tr>
<td>AM-1</td>
<td>8.3</td>
<td>0.201</td>
<td>-</td>
</tr>
<tr>
<td>BI-0.05</td>
<td>0.415</td>
<td>-</td>
<td>0.05</td>
</tr>
<tr>
<td>BI-0.1</td>
<td>0.83</td>
<td>-</td>
<td>0.1</td>
</tr>
<tr>
<td>BI-0.5</td>
<td>4.15</td>
<td>-</td>
<td>0.5</td>
</tr>
<tr>
<td>BI-1</td>
<td>8.3</td>
<td>-</td>
<td>1</td>
</tr>
</tbody>
</table>

Table 1: Code of EMS and Varenets samples
Material and methods

Materials

The cow's skimmed milk (fat 0.05%, Valio, Russia) was used for Varenets. Native and enzyme-modified potato starches (modification by Amylosubtilin® (AM) and Amylase of Bacillus licheniformis (B)) have been described by us earlier13,14. Enzyme preparation of Amylase of Bacillus licheniformis have been described earlier14. Amylosubtilin® (Berdsk Factory of Biological Preparations (now: Sibbiofarm), Russia) is obtained by drying the fermentation broth upon the in-depth cultivation of Bacillus subtilis. These starches (Tabl.1) have been investigated by us earlier.

Production of Varenets

The starches were added to the milk at a concentration of 1% and heated, followed by heating at 98 °C for 3 hours to obtain melted milk, and the mixture of melted milk and starch were mixed for the even distribution of starch. Similarly, a control sample of milk without starch was heated too. Starter culture of lactic acid bacteria (Lactobacillus delbrueckii subsp. bulgaricus and Streptococcus thermophiles) was fermented at 37 °C for 16 hours. The sourdough is sated (5% vol/vol.) into the control milk or milk-starch mixture after cooling at 40 °C. Incubation at 40 °C for 6-7 hours before reaching pH 4.5 and stabilizing at 6 degrees. After stabilization, each Varenets sample was evaluated for physicochemical, textural, sensory, antioxidant properties.

Physicochemical Analyses

The amount of protein, lactose (carbohydrates), content of solid substances and density were measured using InfraLUM® FT-12 (Russian Federation) as described earlier13. Proteins, salts in whey were tested on the milk analyzer Clover-2M (Russia). The whey was obtained by centrifugation of samples (at 3000 g 15 min). A pH meter (HI 2216, Hanna Instruments, Germany) was used to measure the pH. The titrative acidity was evaluated by titration of the suspension (10 g yoghurt in 20 ml distilled water). The samples were boiled to remove carbon dioxide and cooled. The sample was titrated with 0.1 M sodium hydroxide (NaOH) to pink in the presence of 1% phenolphthalein as an indicator. Methods for testing the water - saving capacity (WHC)13 and syneresis were described by us earlier. The apparent viscosity of the Varenets (300 ml, at 8 C) was measured with a Brookfield concentric cylinder viscometer (Model RV-DVIII, Inc., China) equipped with rotor No. 3 at 60 rpm. The measurements were carried out in triplicate for each treatment and the results were recorded in mPa • s.

The colour parameters (L*, a*, and b*) of the yogurt samples were measured using a colorimeter Chroma Meter (China).10

Determination of the Total Amount of Polysaccharides in Varenets

The isolation of exopolysaccharides and polysaccharides (total TAP polysaccharides) and their determination were carried out as described by Feldmanel et al. (2014)15 with modifications. 10-15 g of the sample was placed in a laboratory flask and boiled in a water bath at 100°C for 30 minutes. The samples were cooled and centrifuged at 4000 rpm for 30 minutes and 0.7 ml of 85% trichloroacetic acid was added to 4 ml of the supernatants. These mixtures were cooled to 4°C and centrifuged again at 8000 rpm for 10 minutes. The deposition of TAP (1 ml) from the supernatant was carried out using cold ethanol (-20°C, 3 ml). The Samples were kept in a refrigerator for 48 h and then centrifuged (4°C, 8000 g, for 10 min), the precipitate was re-dissolved in distilled water (volume is equal to the sample volumes), and the TAP was determined as described by Feldmanel et al. (2014).15

Sensory Evaluation

Varenets samples were evaluated by panelist as described by Nikitina et al. 2019.13 Sensory assessment was conducted by ten trained participants (5 men, 5 women, age 18-45 years, healthy subjects tolerant to lactose). Hedonistic scale from 1 (very dislike) to 10 (very pleasant) was used for taste, from 1 (poor) to 3 (excellent) for odour, from 1 (poor) to 10 (excellent) for texture and consistency and 1 for evaluation of yogurt appearance. Defects were also requested if found. Each participant received 30 ml of yoghurt (at 12°C) in a 100-ml glass bottle with a twisting lid.

Evaluation of Antioxidant Capacity

1 DPPH (2,2-di-phenyl-1-picryl hydrazyl) Radical Scavenging Assay

Antioxidant scavenging capacity was analyzed according to the procedure described by Brand-Williams et al., 199516, with some modifications. Samples of Varenets and their whey’s were
previously diluted 10 times, after which 1 ml of diluted samples were mixed with 1 ml of a freshly prepared solution DPPH (0.12 mM) in ethanol. Reaction mixtures and control (2 ml; DPPH in ethanol) were incubated at room temperature in the dark for 30 minutes. The mixtures were centrifuged for 2 minutes, 10,000 rpm. The absorbance of the supernatant was measured at $\chi_{\text{max}}$ 517 nm using a spectrophotometer and DPPH radical capture activity was calculated.

**Ferric Reducing Antioxidant Power Assay (Frap)**
The reduction power analysis was carried out according to the procedure described by Lertittikul et al., 2007, with modifications. 1 ml of the test product (whey) was mixed with 1 ml of 0.2 M potassium-sodium phosphate buffer (pH 6.5) and 1 ml of 1% potassium ferricyanide. The reaction mixture was incubated for 20 minutes at 50°C, cooled, and then 1 ml of 10% trichloroacetic acid was added. The mixture was centrifuged at 2000 g for 10 minutes at room temperature. 2 ml of distilled water and 400 μl of 0.1% FeCl3 were added to the supernatant (2 ml). The absorption of the reaction mixture was measured at 700 nm. The reducing force was expressed as absorption at 700 nm relative to the control.

**Statistical Analysis**
All experimental data were obtained from at least three parallel experiments, the results of analytical determinations for each sample in three or five parallel experiments. A simple classification variance analysis was applied, and whenever it was adequate, the Tukey’s test was used to determine differences between samples. Reliability was established at $p \leq 0.05$. Data analysis was performed using GraphPad software.

**Results and Discussion**

**Physicochemical Composition Varenets Sample**
The pH and titratable acidity of the samples of Varenets are shown in figure 1. The pH of the control and native samples was at the level of 4.35–4.4, whereas samples with BI starches had lower pH (4.28–4.32). The lowest pH was in AM samples. Titratable acidity reflects dependence, the lower the pH, the higher the acidity. The exception is sample AM-1. The introduction of enzyme-modified starch obtained using Amylosubtilin stimulates acidity growth during lactose fermentation. Possibly the starch, that has undergone preliminary hydrolysis acts as an additional source of energy.

![Fig. 1: pH measurements and titratable acidity of non-fat Varenets sample with added different starches: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by Amylosubtilin), BI-0.05, BI-0.1, BI-0.5, BI-1 (modification by Amylase of Bacillus licheniformis)](image)

The chemical composition of Varenets is shown in Table 1. In general, there is no significant difference in the total amount of protein in the control Varenets and EMS samples, however the amount of protein in the Native sample is 0.4% less than in the samples of others ($P > 0.05$). The amount of whey protein in the
AM and Bl samples in total increased by about 0.2%, which may be due to the more active development of lactic acid bacteria and the manifestation of their proteolytic activity, which increases the amount of peptides as a product hydrolysis of casein. The amount of carbohydrates and the total dry matter content of the starch samples naturally increased as a result of the addition of 2% starch carbohydrates. The increase in whey salts and beverage density was due to the addition of starch. Other studies have revealed similar trends, increased acidity and decreased pH in fermented milk beverages and yoghurts with the addition of modified corn and cassava starch.\textsuperscript{18,19,20}

Table 2: Physicochemical composition Varenets sample with added different starches: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by Amylosubtilin\textsuperscript{®}), Bl-0.05, Bl-0.1, Bl-0.5, Bl-1 (modification by Amylase of Bacillus licheniformis). The data presented is the average of three replicates with standard deviation.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Protein, %</th>
<th>Whey protein, %</th>
<th>Carbo-hydrates, %</th>
<th>Solids contents, %</th>
<th>Salts, %</th>
<th>Density, kg/m\textsuperscript{3}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control*</td>
<td>3.46±0.23</td>
<td>1.96±0.02\textsuperscript{**}</td>
<td>4.06±0.04\textsuperscript{**}</td>
<td>8.21±0.3\textsuperscript{**}</td>
<td>0.69±0.01\textsuperscript{**}</td>
<td>1033±1\textsuperscript{**}</td>
</tr>
<tr>
<td>Native</td>
<td>3.09±0.21</td>
<td>2.04±0.04</td>
<td>6.09±0.03</td>
<td>10.72±0.29</td>
<td>0.71±0.01</td>
<td>1036±0.5</td>
</tr>
<tr>
<td>AM-0.05</td>
<td>3.50±0.32</td>
<td>2.3±0.04</td>
<td>6.06±0.07</td>
<td>10.87±0.44</td>
<td>0.77±0.01</td>
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</tr>
<tr>
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<td>0.76±0.00</td>
<td>1037±0.1</td>
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<tr>
<td>AM-0.5</td>
<td>3.47±0.24</td>
<td>2.26±0.03</td>
<td>6.06±0.06</td>
<td>10.83±0.34</td>
<td>0.76±0.01</td>
<td>1036±1</td>
</tr>
<tr>
<td>AM-1</td>
<td>3.46±0.26</td>
<td>2.22±0.03</td>
<td>6.05±0.06</td>
<td>10.73±0.36</td>
<td>0.76±0.01</td>
<td>1036±1</td>
</tr>
<tr>
<td>BL-0.05</td>
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<td>2.27±0.10</td>
<td>6.07±0.10</td>
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<td>0.76±0.01</td>
<td>1036±1</td>
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<td>BL-0.1</td>
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<td>2.25±0.04</td>
<td>6.06±0.10</td>
<td>10.83±0.39</td>
<td>0.76±0.00</td>
<td>1036±0.1</td>
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<tr>
<td>BL-0.5</td>
<td>3.48±0.20</td>
<td>2.28±0.04</td>
<td>6.06±0.08</td>
<td>10.83±0.33</td>
<td>0.76±0.01</td>
<td>1036±1</td>
</tr>
<tr>
<td>BL-1</td>
<td>3.52±0.35</td>
<td>2.27±0.04</td>
<td>6.06±0.08</td>
<td>10.88±0.48</td>
<td>0.76±0.01</td>
<td>1036±1</td>
</tr>
</tbody>
</table>

* (not fat Varenets), ** - indicate significant differences among groups (P ≤ 0.05).

Fig. 2: Syneresis (A) and WHC (B) of non-fat Varenets samples added with different starches: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by Amylosubtilin\textsuperscript{®}), Bl-0.05, Bl-0.1, Bl-0.5, Bl-1 (modification by Amylase of Bacillus licheniformis). The data presented is the average of three replicates with standard deviation.

* - indicate significant differences control group and others (P ≤ 0.05), ** - indicate significant differences native group and others (P ≤ 0.05).
Syneresis and WHC
The increase of the dry residue due to the addition of starch should have a positive effect on the textural characteristics of Varenets. It was found that the introduction of enzyme-modified starches AM-0.5, AM-1, BL-0.1, BL-0.5, BL-1 positively affects the level of syneresis of samples (Figure 2A). The native starch sample had the highest syneresis. A long chain molecule of the native starch could inhibit the close association of casein, and a more open structure could be expected (Schmidt et al., 2001). Such a positive effect of EMC can be due to their lower viscosity, increased water absorption capacity (WAS) and solubility at room temperature (WSI), as well as morphological rearrangements, loosening of starch granules, respectively, an increase in the total contact area with dairy components. The behavior of syneresis observed in this study coincides with that of fermented milk beverages and starch-added yoghurts.

The WHC of the control sample and the EMS samples were at the same level, whereas native starch increased this capacity in the milk clot by 18% (Figure 2B). This appears to be due to the high gelling capacity of native potato starch after heating and, accordingly, the greater ability to retain water in the matrix after heat treatment. Native starch has a high swelling capacity at high temperatures, which provides high WHC.

Apparent Viscosity
The viscosity of the starch-added samples was significantly higher than the control (Fig. 3). The viscosity of Varenets with EMC was higher than that of native Varenets starch, especially when using starches AM-0.5, AM-1, BL-1 in Varenets technology. The mechanism for increasing viscosity and, as a result, the textures of fermented milk beverages varies depending on the degree of hydrolysis of EMS. Some of the BL samples had low viscosity and syneresis, the starch samples had a higher water absorption capacity than native starch. This property provides a better ability to bind water in the syneresis test when the mechanical stress is low.

Fig. 3: Viscosity of non-fat Varenets sample with different potato starches: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by Amylosubtilin®), BL-0.05, BL-0.1, BL-0.5, BL-1 (modification by Amylase of Bacillus licheniformis). The data presented is the average of three replicates with standard deviation.

* - indicate significant differences control group and others (P ≤ 0.05). ** - indicate significant differences native group and others (P ≤ 0.05).

Modified starches are generally neutral stabilizers that do not interact with proteins in pH-dependent processes, but improve the consistency and texture of the beverage by increasing the viscosity of the aqueous phase of the system. Although the viscosity of the native starch gel is higher,
the viscosity of the fermented product with native starch is lower than with EMS, this may be due to the looser and micelle-like structure of the EMS gel in milk and its ability to form more homogeneous matrices. An increase in the concentration of the enzyme used to produce starch makes it possible to obtain starch with a lower molecular weight. This structure of partially hydrolyzed starch seems to allow the starch to react more effectively with milk proteins, especially during a 3-hour heating process. *Amylosubtilin* and *B. licheniformis* amylase have been used for modification in the form of complex preparations having little activity, such as beta-amylase, cellulase, protease, etc. For this reason starches with different structure and characteristics have been obtained.

**Total Amount of Polysaccharides in Varenets**

Exopolysaccharides (EPS) are important metabolites of LAB. They can be emitted into the environment as EPS mucus or adhere to the bacterial surface, forming capsule EPS. EPS dairy products typically act as natural and safe thickeners, emulsifiers or stabilizers, resulting in a significant improvement in texture. In the case of skimmed dairy products, natural EPS may not be sufficient to create the desired texture. The addition of starch to the Russian defatted Varenets resulted in an increase in extractable polysaccharides, the difference between the control and the native sample was 16 mg/g of product (Fig. 4). For EMS samples, the increase in TAP total concentration was 20-25 mg/g of the product. The largest amount of TAP was found in all BI and AM-1 samples. Apparently, EMS appears to act as an additional substrate or stimulating agent for the synthesis of polysaccharides by lactic acid bacteria.

Increased synthesis of EPS positively affects viscosity and syneresis, EMS further hydrophilizes casein aggregates. As a result the probability of phase division at which the size of the areas of the dispersive environment reducing sample durability decreased.

![Fig. 4: TAP of non-fat Varenets sample with different starches: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by *Amylosubtilin*®), BI-0.05, BI-0.1, BI-0.5, BI-1 (modification by Amylase of *Bacillus licheniformis*). The data presented is the average of three replicates with standard deviation. * - indicate significant differences control group and others (P ≤ 0.05).](image)

**Effect of Ems on Sensory Characteristics and Color**

Table 3 shows the results of the study of the sensory properties of Varenets (taste, appearance, texture, aroma, and in general). Most dairy consumers are aware of the positive effects of low-fat yogurt, but the texture and taste are not always satisfactory. In this regard, low-fat or fat-free dairy products are less popular among consumers due to the loss of rheological and sensory properties, as well as an...
increase in syneresis. In the present study, EMS had a positive effect on the sensory evaluation of fat-free Varenets samples. The native starch sample was the least preferred in appearance; the remaining starch samples did not differ from the control. When the aroma was tested, the most intense pleasant smell of heating in combination with fermented milk was in samples AM-0.1 and BI-1, in the native sample the aroma was unimpressed, without an intense taste of lactic acid. When evaluating the texture, it was found that the smoothest, homogeneous and viscous consistency is characteristic of samples AM-0.1, AM-0.5, as well as B1-0.5, B1-1.

The control sample was less viscous and viscous, with a less pronounced heating aroma. The average taste score of samples with EMS was higher than that of the control sample; Varenets AM-0.1, AM-0.5 and BI-0.5, BI-1 had high taste qualities. The native sample had a taste of starch and was poorly lubricated on the palate, since it had a ragged consistency. During heating milk, the melanoidin formation reaction occurs, as a result of which the milk acquires a cream tint and a specific taste. After fermentation with lactic acid bacteria, the taste intensity of the Maillard reaction products decreased due to acidic taste.

Lactic fermentation did not affect color characteristics, but the presence of starch slightly changed the color characteristics of Varenets (Table 3). Compared to the control sample, Varenets with starch was darker, the native sample had the greatest darkening, enzymatically modified starches did not lead to such intense darkening. Studies of starch thermal stability have shown higher resistance to EMS than native starch (data not shown). This may cause more intense staining of native starch Varenets because a 3-hour heating period resulted in partial decomposition of native starch and release of glucose to form reactive milk proteins. In AM and B1 samples, indices a and b are more shifted to the red and yellow sides, respectively, than in the control sample. The Maillard reaction when EMS is added to milk is less intense than when native starch is added. From the point of view of the technological process, in order to prevent a more intense color of the Varenets, it is possible to reduce the heating time, which will reduce economic costs.

**Table 3: Sensory scores and color measurements of non-fat Varenets sample added with different EMS: Control (without starch), Native (potato starch), AM-0.05, AM-0.1, AM-0.5, AM-1 (modification by Amylosubtilin®), BI-0.05, BI-0.1, BI-0.5, BI-1 (modification by Amylase of Bacillus licheniformis)**

<table>
<thead>
<tr>
<th>Sample</th>
<th>Appearance (0-1)</th>
<th>Odour (1-3)</th>
<th>Texture (1-10)</th>
<th>Taste (1-10)</th>
<th>Overall</th>
<th>Color</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>L1</td>
<td>a1</td>
<td>b1</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Control</td>
<td>1±0</td>
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<td>5.8±0.8</td>
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<td>13.8±1.9</td>
<td>71.23±1.9</td>
</tr>
<tr>
<td>Native</td>
<td>0±0</td>
<td>1.0±0.7</td>
<td>4.3±0.6</td>
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<td>9.6±1.9</td>
<td>52.77±2.11*</td>
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<td>7.2±0.5</td>
<td>6.8±0.5</td>
<td>16.8±1.3</td>
<td>66.38±4.21</td>
</tr>
<tr>
<td>BL-0.05</td>
<td>1±0</td>
<td>1.6±0.6</td>
<td>6.8±0.5</td>
<td>7±0</td>
<td>16.4±1</td>
<td>67.69±3.98</td>
</tr>
<tr>
<td>BL-0.1</td>
<td>1±0</td>
<td>1.6±0.6</td>
<td>7.8±0.5</td>
<td>7.2±0.8</td>
<td>17±1.8</td>
<td>64.30±1.98</td>
</tr>
<tr>
<td>BL-0.5</td>
<td>1±0</td>
<td>1.8±0.5</td>
<td>8.2±0.5</td>
<td>7.2±0.5</td>
<td>18.2±1.3</td>
<td>65.88±2.12</td>
</tr>
<tr>
<td>BL-1</td>
<td>1±0</td>
<td>2±0</td>
<td>8.6±0.6</td>
<td>8.2±0.5</td>
<td>19.8±1.0</td>
<td>68.62±2.14</td>
</tr>
</tbody>
</table>

*L, darkness-lightness (0–100); a, greenness-redness (−60 – +60); b, blueness-yellowness (−60 – +60).
* - indicate significant differences control group and others (P ≤ 0.05).
samples had the greatest potential for binding free radicals (Fig. 5A); in these samples, the serum also had high DPPH-activity. The Native sample showed the least activity. Whey of AM samples in 3 out of 4 cases and native whey showed comparable DPPH activity with control.

The ability to reduce Fe$^{3+}$ in Fe$^{2+}$ was higher than that of the fermented milk product with EMS (Fig. 5B), while the control and native showed weaker strength. The addition of starches to Varenets resulted in an increase in the regenerative capacity of the product whey. The greatest reducing potential was shown by BL Varenets starch whey.

The high antioxidant potential of lactic acid bacteria has been reported in many sources, the ability to bind peroxides and free radicals is found not only in the supernatant of the culture fluid, cell-free extracts, but also in fermented dairy products, individual protein components isolated from dairy products. However, studies of the role of starch in the formation of the antioxidant potential of lactic acid products are not enough. Wu et al. described that raw potato starch hydrolysates have high antioxidant activity and can be used as a food antioxidant. In our case, hydrolysis products do not play a significant role in the formation of antioxidant potential, since they are present in starch in a residual amount.

**Conclusion**

The use of enzyme-modified potato starches to stabilize skimmed Varenets results in a higher protein level than native starch, while increasing whey protein levels. In particular, EMS, especially those treated with amylase B. licheniformis added to skimmed milk, do not interfere with the starting culture of Lactobacillus delbrueckii subsp. bulgaricus strain, and Streptococcus thermophilus fermenting lactose to lactic acid in the production of Varenets.

Starches do not have significant antioxidant potential, but partially fermented starch, as a representative of the low molecular weight polysaccharide in the composition of Varenets, stimulated the accumulation of lactic acid (Fig. 1), the accumulation of exopolysaccharides (Fig. 4). That is, such starch can be considered as an additional carbon source that contributes to the development of starters and the formation by lactic acid bacteria of a complex of substances with antioxidant properties.
stimulate the synthesis of exopolysaccharides. Varenets with EMC has a lower level of syneresis than the native and control sample, also due to enhanced EPS synthesis.

Under the influence of EMC, the viscosity of Varenets increases, the texture of the drink improves, and the ability of the milk gel to bind whey improves. EMS at a concentration of 1% can be added to the Varenets mixture to produce a more creamy taste of melted defatted yogurt products from baked milk. Varenets prepared from EMS had a higher level of antioxidants than that prepared from native starch. Varenets samples with BL starch showed the highest antioxidant capacity for DPPH and FRAP tests. Such starches contribute to the formation of a greater antioxidant potential in the final fermented milk product, which increases the utility of the defatted stabilized Varenets. In addition, in vivo studies are needed to assess the possible health benefits of a Varenets dairy product with EMS, such as free radical damage protection.

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**Conflict of Interest**
The authors declares no conflict of interest.

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